

GUSTATORY EFFECTS ON INTESTINAL MOTILITY OF DOGS

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Summary : Trained, unanaesthetised dogs with jejunal fistula and adapted to 2 h meal-time showed transient taste-correlated changes in pressure (mm H₂O) but not in frequency of intestinal motility. Intestinal pressure was increased on bitter taste both before meal-time (4.7 ± 0.2 mm) and after it (13.1 ± 0.9 mm) over respective basal pressure (before meal 3.2 ± 0.4 mm, after meal 10.6 ± 1.4 mm), whereas it was decreased on sweetness of saccharin (before meal 1.1 ± 0.1 mm, after meal 4.8 ± 0.5 mm), and after glucose (before meal 1.7 ± 0.2 mm; after meal 8.8 ± 0.9 mm). Taste-induced motility changes were more pronounced on starvation than on fed state.

Key words : intestinal motility

taste

INTRODUCTION

The gastro-intestinal secretions and motility both of which are essential for digestion, absorption and movement of food in the gastro-intestinal tract, are known to be increased in the presence of food in the stomach (1). Further it is also known that before food actually enters the stomach, its fast-acting sensory properties (sight, smell, taste etc.) enhance salivary, gastric (7) and pancreatic secretions (5) thus acting as primers to secretions after the arrival of food. But such anticipatory effects of sensory cues on clinically more important intestinal motility (3) are virtually unknown and hence the present report.

MATERIAL AND METHODS

Trained young (6-8 months old) 2 h meal-time (1300-1500 h) adapted and conscious dogs (n=6) were used for the investigation. Their intestinal motility responses to taste both before and after meal-time were recorded kymographically using conventional balloon method (2) and water manometer as described by us earlier (10). From a pipette

with a fairly long (3 cm) nozzle 2-3 ml of any one of gustatory test solutions (18% glucose, 0.8% saccharin and 0.006% quinine in distilled water at room temperature of 24-26°C) on anyone test day was dropped (from 1.0-1.5 cm height) on tongue *in situ* of the dog lying quietly on a table and gently restrained.

RESULTS

Taste effects on the on-going (basal) intestinal motility were seen 20-40 sec after placement of test solution and for a period of 30-100 sec, both before and after food (Fig.1). The visually observed smacking, tongue-rolling and swallowing movements

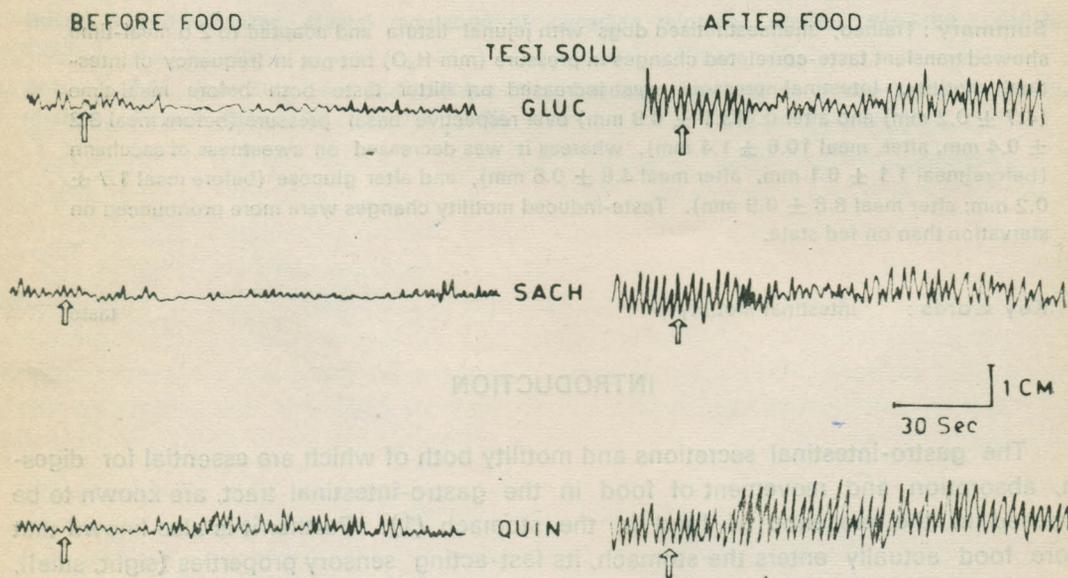


Fig. 1 : Effects of gustation on intestinal motility of fed (after food) and starved (before food) dogs. Arrow (↑) mark indicates the point in time at which test solution was placed on tongue.

which started almost immediately (5-10 sec) and lasted for nearly 1 min period, following test solution contact with tongue, had no influence on intestinal motility. The sweet taste of a substance whether containing calories (glucose) or not (saccharin) decreased the intra-luminal pressure (mm H₂O) of on-going basal intestinal activity whereas bitter taste (quinine) increased it. The taste (either sweet or bitter) did not affect the frequency (waves/min) of motility, and in that respect is similar to effects after food intake which also did not alter frequency though it caused significant increase in intra-luminal pressure (Table I). The effects of taste on low-amplitude starvation (before food) - correlated,

basal motility were more pronounced, both in force and duration (Fig. 2), than on after-food-induced high amplitude basal motility.

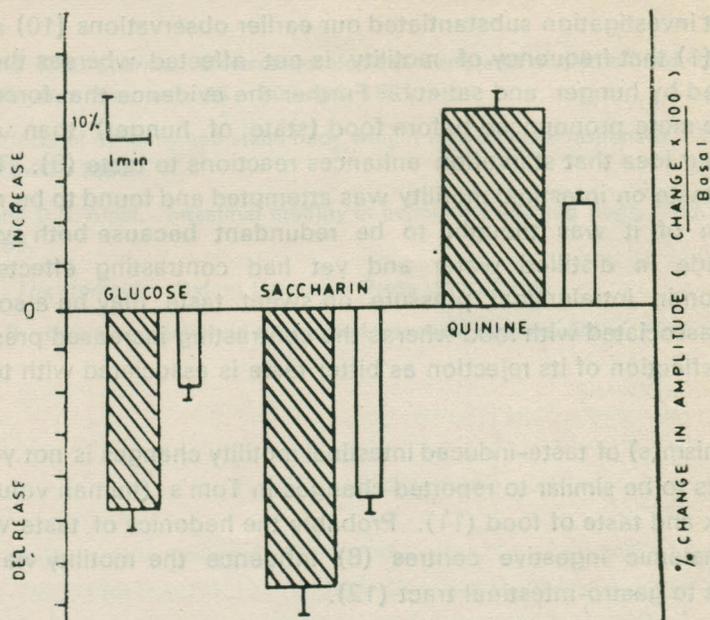


Fig. 2 : Effects of gustation on amplitude (vertical height of bar), duration (horizontal length) and direction (increase or decrease) of dogs before (■) and after (□) food.

TABLE I : Effects of gustation on intestinal motility.
(mean \pm SE) of dog.

	BASAL	GLUCOSE	SACCHARIN	QUININE
1. Before food				
Pressure (mm H ₂ O)	3.2 \pm 0.4	1.7 \pm 0.2*	1.1 \pm 0.1*	4.7 \pm 0.2*
Freq. (Waves/min)	25.8 \pm 1.1	23.8 \pm 0.9	21.8 \pm 1.2	27.3 \pm 1.1
2. After food				
Pressure (mm H ₂ O)	10.6 \pm 1.4	8.8 \pm 0.9	4.8 \pm 0.5*	13.1 \pm 0.9*
Freq. (Waves/min)	33.4 \pm 1.7	31.5 \pm 1.5	30.4 \pm 1.8	34.8 \pm 1.2

*P < 0.05 compared to basal

DISCUSSION

The present investigation substantiated our earlier observations (10) and the observations of others (1) that frequency of motility is not affected whereas the intra-luminal pressure is altered by hunger and satiety. Further the evidence that force and duration of taste effects are more pronounced before food (state of hunger) than after food (fed state) reinforced our idea that starvation enhances reactions to taste (9). Though effects of distilled water taste on intestinal motility was attempted and found to be not significant, a specific mention of it was thought to be redundant because both sweet and bitter solutions were made in distilled water and yet had contrasting effects on intestinal motility. Reduction in intraluminal pressure on sweet taste may be a sort of receptive relaxation as it is associated with food whereas the contrasting increased pressure on bitter taste could be a reflection of its rejection as bitter taste is associated with toxicity (4).

The mechanism(s) of taste-induced intestinal motility changes is not yet worked out. However it appears to be similar to reported changes in Tom's (human volunteer) gastric motility on the taste and taste of food (11). Probably the hedonics of taste with its known effects on hypothalamic ingestive centres (8) influence the motility via hypothalamic neural connections to gastro-intestinal tract (12).

The present report indicates that taste, like the peptide motilin, exerting its "primary effect on gastro-intestinal motility can have a profound effect on metabolism" (6) through the effects on intestinal transit time and absorption.

REFERENCES

1. Davenport, H.W. In : *Physiology of digestive tract*. Year Book of Medical Publications Inc, Chicago, 1982.
2. High Tower Jr, N.C. Motor action of small bowel. In : *Hand book of Physiology Sec. 6. Alimentary Canal. IV* (Ed) C.F. Code American Physiological Society, Washington, DC, 1967.
3. High Tower Jr, N. C. and H. D. Janowitz. In : *Physiological basis of Medical Practice* (Ed) J.R. Brobeck, Williams and Wilkins Co, Baltimore, 1979.
4. Le Magnen, J. Habits and Food intake. In : *Hand book of Physiology, Sec 6: Alimentary Canal 1*, (Ed) C.F. Code, American Physiological Society, Washington, DC, 1967.
5. Naim, M. and M.R. Kare. Taste stimuli and Pancreatic functions. In ; *Chemical senses and Nutrition*, (Ed) M.R. Kare and Owen Maller, Academic Press, New York, 1977.

6. Nicholl, C. G., J.M. Polak and S. R. Bloom. Hormonal regulation of food intake, digestion and absorption. *Ann. Rev. Nutr.*, **5** : 213-239, 1985.
7. Pavlov, I. P. *The work of digestive glands*. (Translated by W.H. Thomson) 2nd Ed, Griffin, London, 1910.
8. Rao, B.S. and K.N. Sharma. Electrophysiological correlates of diurnal feeding rhythm disruption in rat. *Ind. J. Physiol. Pharmac.*, **25** : 308-318, 1981.
9. Rao, B.S. Effects of dynamic and static body weight loss on taste responses. *Ind. J. Physiol. Pharmac.*, **27** : 25-31, 1983.
10. Rao, B. S. and D.S. Amar. Intestinal motility in helminthic infested dogs. *Ind. J. Exptl. Biol.*, **23** : 279-280, 1985.
11. Wolf, S. In : *The stomach*. Oxford University Press Inc., 1965.
12. Youmans, W.B. Neural regulation of gastric and intestinal motility *Am. J. Med.*, **13** : 209-226, 1952.

INTRODUCTION

An apparatus for measuring reaction time to both visual and auditory stimuli with a provision to vary intensity of stimuli and with facility for lateralisation of sound to the right or left ear has been manufactured. The instrument is being used for the study of reaction time in normal, smokers, diabetic, psychiatric patients and neurologically affected patients.

Description of the reaction time assembly: The electronic apparatus to measure reaction time has an eight digit display unit for time interval measurement and intensity controls for visual stimulus by energising an electric lamp (fitted at rear side of the equipment) at the set intensity or alternatively for auditory stimulus by energising an audio amplifier at the set intensity. The sound output is either through a loud speaker or through a stereo head phone. (Fig. 1) When the headphone is connected, the sound can be heard either through the right left or both ears by selecting the switch position marked as R, L or B provided on the rear of the amplifier. Accurate resolution in time interval measurement is 100 or 10000000.

The time circuit is constructed using standard I.C.s and modulation IC type ICM 7228 A manufactured by Intel, Inc. U.S.A. The ICM 7228 A can function as frequency counter, period counter, frequency ratio counter or time interval counter. The block diagram of the entire assembly is shown in Fig. 2.